The role of regulatory T cells in the regulation of upper airway inflammation

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ABSTRACT

Allergic rhinitis (AR) and chronic rhinosinusitis with nasal polyps (CRSwNP) are inflammatory diseases of the upper airway, with a similar immunologic profile, characterized by aberrant and persistent type 2 inflammation. One cell population that has been identified as altered in both disease types is regulatory T cell (Treg). Tregs have the capacity to modulate T-effector function and suppress inflammatory cytokine production in a broad range of cell types. Given the ability of Tregs to control inflammation, the role of Tregs in respiratory diseases has attracted much attention. As discussed in this article, alterations in the Treg numbers and function, or both, have been identified in AR and CRSwNP, although much of the data is conflicting. Here, we explored what is known and, in many cases, unknown about the mechanisms by which Tregs differentiate and function, and how these functions can be controlled in the mucosal microenvironment. By gaining a greater understanding of these processes, it may be possible to harness the natural immunosuppressive activity of Tregs to ameliorate the chronic inflammation associated with AR and CRSwNP.

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INCIDENCE AND HEALTH CARE BURDEN OF AR AND CRS

AR has consistently increased in prevalence and now affects up to 6% worldwide 2 and up to 40% of adults in the United States. 3 With symptoms regularly increasing or decreasing, its severity varies constantly, which makes it difficult to gauge. 2 As such, AR can have a profound effect on quality of life, especially in children. Although it can be difficult to quantify, the annual cost of AR was estimated in 2004 to be between $2 billion and $5 billion/y in the United States. 4 CRS affects up to 12% of the U.S. population, with direct and indirect costs that surpass $21 billion/y in the United States. 5 CRS is a complex disease, with two different presentations, CRS with nasal polyps (CRSwNP) and without nasal polyps (CRSsNP).

CRSsNP can be caused by a broad range of disorders, including structural abnormalities (e.g., nasal septal deviation, septal perforation, nasal valve dysfunctions) and other pathologies (e.g., primary ciliary dyskinesia or gastroesophageal reflux). 6 CRSNP has historically been connected with T-helper (Th) type 1 skewing, although results of some studies indicated a wide variation in the immune profile expressed, 7,8 with reports of tissue immunologic characteristics even seeming similar to those of controls. 3 CRSwNP, in addition to the polyps themselves, presents with a characteristic type 2 eosinophil-skewed immune response, similar to that seen in AR. The close relationship between AR and CRSwNP may be, in part, due to the similar type 2 skewed immune profile and the fact that 82% of CRSwNP are atopic. 9 Therefore, CRSwNP was the primary focus of this article.

One immune cell subtype, Treg, has experienced an increase in interest and relevance as technology has progressed to the point that we can accurately study it. Once known as suppressive T cells in the 1980s, Tregs have been established as a powerful force in the immunologic milieu, with far reaching and potent effects on self-tolerance, prevention of autoimmune disease, and allergy. 10 The purpose of this review was to examine the current state of knowledge on the role of Tregs in AR and CRSwNP.

Treg DEVELOPMENT AND FUNCTION

One of the first hurdles to understanding the role of Tregs in disease was to establish a set of markers by which to define Tregs, which remains a problem today because no marker unique to Tregs alone has been found. However, as with other T-cell phenotypes, a combination of markers has been used to accurately identify Tregs as a unique population of cells. Tregs are commonly defined as CD4+ CD25+ CD69+ and express CD127low/-, and have been found to have a maximum suppressive ability with the CD45RA-CD45RO- phenotype. 10,11 However, not all Tregs are CD4+. Subsets of highly suppressive CD8+ CD25+ Forkhead Box P3+ (FoxP3+) Tregs in both humans and mice have been identified. 12-14 Although CD8+ Tregs have been identified in the pathophysiology of other diseases, there is limited information about their role in respiratory diseases. As such, this review focused discussions primarily on CD4+ Tregs.

Tregs also express, on their surface, the regulatory mediator cytotoxic T-lymphocyte-associated protein 4 (CTLA-4). One of the most important markers that has been used for their identification is the FoxP3 transcription factor, which is integral to suppressive activity, differentiation, and function. 15 Although FoxP3 is one of the most widely used markers for Tregs, it is also expressed by non-Tregs when induced to do so, and evidence has even been found of FoxP3 expression at the double-positive stage of T-cell differentiation. 16 Because this was not discovered until relatively recently, older studies on the function and presence of Tregs that solely used FoxP3 to identify them may have been measuring activated effector T cells in addition to Tregs and, therefore, may be inaccurate. Further investi-
gations have led to the discovery of two unique subgroups of Tregs, their thymic Tregs (tTreg) (in older literature, referred to as natural T regulatory cell [nTreg]) and Tregs induced in the periphery (pTreg) (in older literature, referred to as inducible T regulatory cell [iTreg]).

Although tTreg formation is not yet fully understood, it is known that tTregs are differentiated within the thymus through a series of interactions with major histocompatibility complex (MHC) self peptides. However, pTregs are converted peripherally from other CD4+ T cells through expression of transforming growth factor beta (TGF-β) and the CNS1 gene. Although not without controversy, there is some evidence that tTregs can be delineated from pTregs by the presence of Helios protein and, in murine models, the extracellular marker neuropilin-1.20,21 FoxP3+ Helios– cells can be identified specifically as tTregs, which do not produce as many cytokines but which are important to the creation of pTregs, whereas FoxP3– Helios– Tregs are pTregs and have greater cytokine expression, especially of interleukin (IL)-10.22 Although the role of Helios has been questioned due to its transient expression in all types of developing T cells,23 pTregs seem to consistently exert more of a regulatory effect through the use of IL-10 even while displaying subset heterogeneity, such as expressing more or less FoxP3.24 Treg lineages can be broken down even further, e.g., CD45RA Tregs tend to use TGF-β for their regulatory effects, whereas CD45RA+ Tregs use CTLA-4 and are more dependent on IL-10.25

Functionality of Tregs can be defined as the ability to suppress T effector and helper cells through cytokine production, induction of apoptosis (via perforin and granzyme B secretion), and blockade of costimulation via CTLA-4. Their functionality is further mediated by the expression level of FoxP3 by the specific Treg.26 Expression of FoxP3 is augmented by Treg T cell receptor (TCR) interaction with MHC-II signals, and deficiency in FoxP3 results in an absence of CD4+CD25+ cells.27 In addition, suppressive functionality may be regulated by expression of Helios protein, which has been found to require intrinsic FoxP3 expression as well as TGF-β signaling, which further highlights the role of FoxP3 as the “master regulator.”

Tregs secrete TGF-β, IL-10, and IL-35 as their main regulatory cytokines. These factors induce a number of functional alterations to other immune cells with which they interact. Typically, IL-10 is one of the primary regulatory cytokines of the immune system, and downregulates all T-cell cytokine expression and macrophage activity as well as enhances B-cell survival. TGF-β has a variety of effects systemically, but, in the context of local immune cell function, it functions similarly to IL-10 and, therefore, inhibits T- and B-cell proliferation. IL-35 is a recently identified cytokine that seems to have anti-inflammatory and immunosuppressive capabilities similar to TGF-β. It suppresses IL-4 and transcription factor GATA-3 to reduce Th2 proliferation.28 A discrepancy between the two cytokines lies in the fact that although TGF-β can be found in many tissues at all times, IL-35 (as well as IL-10) are not typically constitutively expressed, except in Tregs.29–30

Treg effector functions are predominantly exerted in the local microenvironment, and thus the migration of Tregs is critical for their important functions. Treg migration is moderated by C-C chemokine receptors (CCR) 4 and CCR8, which respond to chemokine (C-C motif) ligand (CCL) 22, CCL17, CCL1, and viral macrophage inflammatory protein-1 (MIP-1).31 Tregs also express C62L, CCR7, and CXCR4, which are indicators of secondary lymphoid homing capacity.32 As summarized in Fig. 1 A, epithelial cells serve as a source of CCL1 and CCL17, whereas CCL22 and IL-16 come from macrophages and Th1 cells, respectively.33–37 IL-16 seems to be a potent chemoattractant, which induces Tregs to migrate and express increased FoxP3 activity. IL-16 also seems to attract Th1 cells, with a net effect in the

Figure 1. The influence of airway-derived factors in regulatory T cell (Treg) migration and differentiation. (A) The presence of Tregs in the airway can be influenced by a number of locally produced mediators. Airway epithelial cells can produce factors such as 1,25-dihydroxyvitamin D3 (1,25(OH)2D3) and transforming growth factor (TGF)-β both of which can induce the differentiation of naive T cells to become Tregs. Epithelial cells also produce chemokatic factors, such as chemokine (C-C motif) ligand (CCL) 1 and CCL17, which induce Treg migration through C-C Chemokine Receptor (CCR) 4 and CCR8, receptors signaling. Mediators made by local immune infiltrates, such as macrophage production of CCL22, and T-helper (Th) type 1 cell production of interleukin (IL) 16 can also induce Treg migration to the airway. (B) Airway-produced factors can also impair Treg functions. Th2 and proinflammatory cytokines, such as IL-4 and tumor necrosis factor (TNF) α, respectively, can skew naive T cells away from Treg differentiation. Impaired epithelial membrane and cell-cell functional integrity in asthma, Allergic rhinitis (AR) and chronic rhinosinusitis with nasal polyps (CRSwNP) allow for the increased passage of antigen into the subepithelial space, which, in turn, can activate dendritic cells and drive them to promote a proinflammatory state characterized by reduced indoleamine 2,3-dioxygenase (IDO).
tissue of Treg-mediated Th2 suppression and normalization of the immunologic population of the tissue.28 Histamine 2 and 4 receptors have been shown to release IL-16, and ligation of the histamine 4 receptor (H4R) receptor results in recruitment of pTregs, which helps to resolve airway hyperresponsiveness and inflammation.29

ALTERATIONS IN Treg NUMBERS AND FUNCTIONS IN AR AND CRSwNP

From studies performed with FoxP3-deficient scurvy mice and observations of immunodysregulation, polyendocrinopathy, enteropathy, and X-linked syndrome in humans with dysfunctional FoxP3, we know that a complete lack of Treg function will lead to significant autoimmune disorders and, ultimately, death.40,41 Use of depletion of Tregs mice has provided conflicting reports about the role of Tregs in lower airway allergic inflammation.42 In studies by Baru et al.,43 which used Balb/c and C57BL/6 mice, which are more resistant to allergy development, Treg depletion during allergen provocation failed to alter lung inflammation. Conversely, depletion of Tregs during the sensitization phase of allergy development resulted in a dramatic exacerbation of inflammation for both strains of mice, which suggests that Tregs may play an important role in the establishment of allergic airway disease.43 To our knowledge, no study has reported the use of depletion of Tregs mice or other Treg depletion methods to assess the role of Tregs in murine models of AR or CRS.

As summarized in Table 1, in patients with AR, circulating Treg numbers are lower than or equivalent in control patients.44–46 Analysis of transcription factor expression by using immunohistochemical techniques conflict with some studies that demonstrated FoxP3, T-box transcription factor (T-bet), and TGF-β expression to be lower in patients with AR than in controls with increased GATA-3 expression,47,48 whereas another study found that FoxP3 and GATA-3 have elevated expression in sinonasal tissue of Treg-mediated Th2 suppression and normalization of the tissue.38 Histamine 2 and 4 receptors and Tregs in peripheral blood and sinus tissue, no difference in circulating levels of either Tregs populations was observed. However, in sinonasal tissue CD4+ CD25+ FoxP3+ Tregs were increased compared with tissue from control subjects. Conversely, CD8+CD25+FoxP3+ Tregs were decreased compared with control subject–derived tissue. In comparisons of patients with CRSwNP who were atopic versus nonatopic, the presence of allergy was found to have no impact on the level of either Treg subset.54 Currently, there is no satisfactory explanation for the differences in findings between the groups. Discrepancies between the studies could be due to different sampling methods or different presentations of the disease; studies found significant variation in cytokine profiles between different countries of origin or ethnicities in CRS.55 Therefore, similar to AR, additional investigation is needed, focused more specifically on Treg function and less on expression of markers to determine their presence or absence.

POTENTIAL CAUSES OF Treg DYSFUNCTION

Changes in the local inflammatory milieu may contribute to altered Treg numbers and/or function in the airway. For example, local upregulation of IL-6 can transform naive T cells into Th17 cells and thus shift away from Treg induction.56 As highlighted in Fig. 1B, the inflammatory cytokine tumor necrosis factor (TNF) α has also been shown to decrease FoxP3 expression and to suppress Treg functions.57 Furthermore, in a murine allergic airway inflammation model, it was demonstrated that TNF-α receptor antagonist reduces inflammation and increases the percentage of Tregs in circulation and their suppressive ability as measured by IL-10 production.58 These findings may have applicability to the upper airway because both patients with AR and patients with CRSwNP have been shown to have increased TNF-α levels.59–61 As mentioned previously, TGF-β promotes naïve T cells to become Tregs. However, patients with AR have a lowered level of TGF-β compared with controls.62 Similarly, patients with CRSwNP have decreased TGF-β in their airways compared with controls.63–65 Loss of this well-known differentiator of naïve T cells into Tregs may contribute to their loss in AR and CRSwNP. However, given that Tregs secrete TGF-β to induce suppression, it is also possible that the loss of TGF-β in AR and may impact the reduction in Treg directly CRSwNP. Therefore, additional studies are needed to extrapolate the exact interplay between TGF-β and Tregs in the upper airway.

Reduced Treg chemotaxis to the tissue from the periphery represents another potential mechanism that contributes to local Treg dysfunction. In patients with allergic asthma, a lack of responsiveness to chemokines is thought to be one potential mechanism that contributes to a loss of local tissue control of immune responses. In patients with allergic asthma, Tregs were found to have decreased chemotactic responses to the Treg chemotactic factor CCL1.66 The mechanisms associated with the loss of chemotactic activity were not elucidated. Whether due to motility issues and/or the loss of chemokine receptor expression is not clear, but the mechanisms that drive these changes warrant a more thorough investigation. Similarly, it was found that chemotaxis of Tregs to mucosal tissues is impaired in CRSwNP;67 however, the precise mechanism responsible for this reduction is yet to be determined. Treg migration, both in airway homeostasis and inflammation, represents an area in which much additional research is needed.

Airway epithelial cell barrier dysfunction is hypothesized to be another contributor to pathogenesis of AR and CRSwNP.70–72 A breakdown in epithelial barrier junctional proteins has been shown to result in a leakiness that facilitates increased passage of antigen, which could hyperstimulate immune cells, particularly antigen-presenting dendritic cells (DC), in the subepithelial space. In AR and CRSwNP, DC numbers are elevated in the local tissues and, in some cases, systemically.73–75 After allergen exposure, these activated and matured DCs stimulate an inflammatory effector T-cell response instead of the tolerogenic phenotype. For example,68 DCs from patients with asthma who were exposed to house-dust mite antigen had reduced indoleamine 2, 3-dioxygenase expression, which shifted the T-cell response toward Th2 and away from Treg induction.76 Inhalation of Aspergillus fumagatis is another potential driver of Treg dysfunction in airway disease. A. fumagatis is a ubiquitous fungal antigen and the most common fungus in the airway, inhaled at a rate of several hundred conidia per day.76–78 Although studies in mice found that a single exposure to A. fumagatis induced increases in Treg numbers and function in the lung, repeated exposure models, more akin to the exposure humans experience, was found to shift Th2 responses toward a pathogenic Th2–Th17 phenotype.79

The secosteroid hormone 1,25-dihydroxyvitamin D3 (1,25[OH]2D3) has been shown in multiple disease states to induce Tregs.80–83 However, patients with CRSwNP have reduced sinonasal levels of 1,25(OH)2D3 compared with controls or those with CRSsNP.84 This deficiency is likely caused by an impaired ability of human sinonasal epithelial cells to metabolize 25(OH)D3 to its activate metabolite, 1,25(OH)2D3.85 These local deficiencies are independent of atopic status, age, race, gender, or vitamin D3 sufficiency status. Therefore, it is possible that local 1,25(OH)2D3 deficiencies may contribute to the reductions in Tregs found in CRSwNP. To date, the local airway impairment of vitamin D metabolism has only been described in CRSwNP, and, therefore, the applicability of these findings to AR remains to be determined.
Table 1  Summary of human studies that examined Treg frequency and functionality in patients with AR or CRSwNP

<table>
<thead>
<tr>
<th>Disease</th>
<th>Change in Tregs vs Control</th>
<th>Sample Source</th>
<th>Treg Markers and Methods</th>
<th>Results</th>
<th>Change in Functionality</th>
<th>Reference No.</th>
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<tr>
<td>AR</td>
<td>↓</td>
<td>PB</td>
<td>CD4+CD25+FoxP3+ expression by flow cytometry</td>
<td>Decreased numbers of circulating Tregs vs controls</td>
<td>TGF-β levels lower in AR than in controls</td>
<td>44</td>
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<tr>
<td></td>
<td></td>
<td></td>
<td></td>
<td>Equivalent numbers of Tregs vs healthy controls</td>
<td>Functionality not impaired</td>
<td>45</td>
</tr>
<tr>
<td></td>
<td>↓</td>
<td>PB</td>
<td>CD4+CD25+FoxP3+ expression by flow cytometry</td>
<td>Higher percentage of CD4+CD25+ T cells but lower FoxP3+ T cells vs controls</td>
<td>IL-10 levels lower in AR vs control, IL-4, IL-5, TNF-α higher</td>
<td>46</td>
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<tr>
<td></td>
<td>↓</td>
<td>NB</td>
<td>FoxP3, GATA-3 and T-bet measured by immunohistochemistry</td>
<td>FoxP3 and T-bet expression was lower in patients with AR vs healthy controls; GATA-3 was the same between the groups</td>
<td>Levels of IL-4, IL-10, TGF-β not different between patients with AR and controls</td>
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<tr>
<td></td>
<td>↑</td>
<td>NB</td>
<td>FoxP3, GATA-3, and T-bet measured by immunohistochemistry</td>
<td>FOXP3+ and GATA-3+ cells were higher in AR vs control preallergy season; in patients with AR levels did not change through the season but declined with fluticasone propionate treatment</td>
<td>Functionality was not measured</td>
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<tr>
<td>CRSwNP</td>
<td>↓</td>
<td>PB</td>
<td>CD4+CD25+FoxP3+ expression by flow cytometry</td>
<td>Lower percentage of circulating Tregs in patients with CRSwNP vs CRSsNP, no difference vs controls</td>
<td>Lower IL-10 and TGF-β and higher TNF-α and IL-6 in CRSwNP vs controls</td>
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<td></td>
<td>↓</td>
<td>SM and IT</td>
<td>T-bet, GATA-3 and FoxP3 mRNA</td>
<td>Decreased FoxP3 expression vs controls; higher T-bet and GATA-3 vs controls</td>
<td>Increased IFN-γ, IL-4, and IL-5, and decreased IL-10 and TGF-β in CRSwNP</td>
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<tr>
<td></td>
<td>↓</td>
<td>SM and IT</td>
<td>FoxP3, T-bet, GATA-3 mRNA; FoxP3 protein by immunohistochemistry</td>
<td>Lower FoxP3 expression in CRSwNP vs controls; higher T-bet and GATA-3 vs controls</td>
<td>TGF-β lower in CRSwNP vs controls and CRSsNP</td>
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<tr>
<td></td>
<td>—</td>
<td>SM and IT</td>
<td>FoxP3 protein by immunohistochemistry</td>
<td>No difference in FoxP3+ cells from IT from controls vs SM samples from patients with CRSwNP</td>
<td>Functionality not measured</td>
<td>53</td>
</tr>
<tr>
<td></td>
<td>- PB, ↓, ↑ SM</td>
<td>PB, SM</td>
<td>CD4 Treg CD3+CD4+CD25+FoxP3+ CD8 Treg CD3+CD8+CD25+FoxP3+ expression by flow cytometry</td>
<td>No difference in PB CD4 Tregs or CD8 Tregs between groups; SM numbers of CD4 Tregs was the same between groups but with lower regulatory potential in CRSwNP mucosa; CD8 Tregs were reduced in CRSwNP SM vs control SM</td>
<td>Lower regulatory potential of CD4+ cells</td>
<td>54</td>
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</table>

Treg = Regulatory T cell; AR = allergic rhinitis; CRSwNP = chronic rhinosinusitis with nasal polyp; ↓ = decrease; PB = peripheral blood; FoxP3+ = FoxP3+ T cells; CD25+ = CD25+ T cells; CD4+ = CD4+ T cells; FoxP3 = FoxP3 protein; GATA-3 = GATA-3 mRNA; mRNA = messenger RNA; ↓ = increase; CRSsNP = chronic rhinosinusitis without nasal polyp.
HARNESSING Tregs FOR THE TREATMENT OF AR AND CRSwNP

Results of studies found that the use of corticosteroids increases both the number and the suppressive function of Tregs in the airways, via an IL-10–dependent mechanism, which suggests a possible mechanism for longer-term resolution of inflammation than just the corticosteroid itself. Allergen-specific immunotherapy has also been proven to increase the number of circulating Tregs; however, this increase is only seen while the immunotherapy is ongoing, with Treg levels dropping to pretreatment levels when allergen exposure ceases. In addition to standard therapy, novel therapeutic agents in humans have the potential to normalize the immunologic profile of the airway. Tregs can be isolated from the blood, expanded ex vivo, and reinfused into the bloodstream. The reinfused cells could be detected circulating for 2 weeks, with no extra risk of infection or early mortality. The results of the application of ex vivo Treg expansion have been promising, but work has primarily been performed in the realms of autoimmune disease, organ transplantation, and graft-versus-host disease.

To our knowledge, there currently are no published studies that have looked at the effect of applying Tregs, especially pTregs, locally to the respiratory mucosa to determine whether it would resolve inflammatory symptoms. To our knowledge, there also are no studies on the effects of IL-2–mediated Treg induction in the respiratory tract. However, analysis of exciting new data indicates that antigen and rapamycin, a tolerogenic immunomodulator, delivered via synthetic polymeric nanoparticles into the respiratory tract and other areas of the body stimulates local Treg proliferation. This effect is seen even in the presence of agonists, which indicates a powerful tolerogenic affect. Although this experiment has thus far only been displayed in mouse models, it could prove a very compelling treatment option.

CONCLUSION

AR and CRS displayed a similar type 2–skewed immunologic profile that was also characterized by the reduced presence of Tregs, especially pTregs. Whereas results of Toll like receptor (TLR) a number of studies in AR and CRSwNP indicated that Treg numbers are decreased, much remains to be understood about the function of Tregs or, in many cases, dysfunction. Current treatments have been found to be effective, at least in part through indirect modulation of Treg populations. However, these effects seem to be transient, and, therefore, given the body of research presented herein, the development of novel strategies to normalize Treg effector responses, numbers, and migration are needed.

REFERENCES


